AMENDMENTS TO THE SPECIFICATION

Please amend the specification as follows:

Please amend the paragraph beginning at page 52, line 13 of the specification to read as follows:

Purified FPDH obtained as described in Example 14 was denatured in the presence of 8 M urea and then digested with Achromobacter-derived lysyl endopeptidase (product of Wako Pure Chemical Industries) and the peptide fragments obtained were sequenced by the Edman method. Considering the DNA sequences deduced from the amino acid sequences, two PCR primers were synthesized (primer 1: 5'-GGNGCNATHGTNAAYATGGG-3' (SEQ ID NO.: 3), primer 2: 5'-CCDATNGGRTGYTGNGTDAT-3'(SEQ ID NO.: 4).

Please amend the paragraph beginning at page 53, line 4 of the specification to read as follows:

Based on the complementary sequence GGAGCGGCCACATACGAGTGAATGG (primer 3(SEQ ID NO.: 5) of a portion close to the 5' side of the core sequence and the sequence AGACACCATTGCTTGATATTTGCCC (primer 4 (SEQ ID NO.: 6) of a portion close to the 3' side, two PCR primers (primer 3(SEQ ID NO.:5) and primer 4(SEQ ID NO.: 6) identical in sequence to those sequences were synthesized.

Please amend the paragraph beginning at page 53, line 10 of the specification to read as follows:

For preparing a template for inverse PCR, the chromosomal DNA of Candida maris IFO 10003 was first digested with the restriction enzyme PstI and the digest was self-circularized using T4 DNA ligase. A buffer solution (100 .mu.l) for ExTaq containing 660 ng of the self-circularization product, the two primers (primer 3, (SEQ ID NO.: 5) and primer 4, (SEQ ID NO.: 6), 100 picomoles each), 20 nanomoles of each dNTP and 2.5 U of ExTaq (product of Takara Shuzo) was prepared, and 40 cycles of thermal denaturation (94 °C., 0.5 min), annealing (55 °C., 0.5 min) and elongation reaction (72 °C., 1 min) were conducted and, after cooling to 4°C., an amplified DNA was confirmed by agarose gel electrophoresis.

Please amend the paragraph beginning at page 54, line 2 of the specification to read as follows:

For causing FPDH expression in Escherichia coli, a recombinant vector to be used for transformation was constructed. First, a double-stranded DNA was prepared which had an NdeI site added to the initiation codon site of the structural gene for FPDH and a new termination codon and an EcoRI site added immediately behind the termination codon, in the following manner. Based on the base sequence determined in Example 21, a primer 5 (5'-CGCCATATGTCCTACAATTTTGCCAAC-3' (SEQ ID NO.: 7)) with an NdeI site added to the initiation codon portion of the structural gene for FPDH and a primer 6 (5'-GCGGAATTCTTATTATCTTGCGGTATAACCACC-3'(SEQ ID NO.: 8)) with a new termination codon and an EcoRI site added immediately behind the termination codon of the structural gene for FPDH were synthesized.

Please amend the paragraph beginning at page 54, line 31 of the specification to read as follows:

A double-stranded DNA comprising the Bacillus megaterium IAM 1030-derived glucose dehydrogenase (hereinafter referred to as GDH) gene with the Escherichia coli Shine-Dalgarno sequence (9 nucleotides) added at 5 bases upstream of the initiation codon of that gene and, further, with an EcoRI digested site added just before that sequence and an SalI digested site added just behind the termination codon was prepared in the following manner. Based on the information on the base sequence of the GDH gene, a primer 7 (5'-GCCGAATTCTAAGGAGGTTAACAATGTATAAAGATTTAGAAGG-3'(SEQ ID NO.: 9)) with the Escherichia coli Shine-Dalgarno sequence (9 nucleotides) added at 5 bases upstream of the initiation codon of the GDH structural gene and, further, with an EcoRI digested site added just before that sequence, and a primer 8 (5'-GCGGTCGACTTATCCGCGTCCTGCTTGG-3'(SEQ ID NO.: 10)) with an SalI site added just behind the termination codon of the GDH structural gene were synthesized in the conventional manner. Using these two primers, a double-

stranded DNA was synthesized by PCR using the plasmid pGDK1 (Eur. J. Biochem., 186, 389 (1989)) as the template. The DNA fragment obtained was digested with EcoRI and SalI, and the digested fragment was inserted into pNTFP constructed in Example 22 at the EcoRI-SalI site thereof to give a recombinant vector, pNTFPG. The construction scheme for and the structure of pNTFPG are shown in FIG. 2.